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Commentary

Signal transducer and activator of transcription 3 (Stat3) is a transcription factor that is constitutively activated in a variety of human malignancies, including prostate, lung, brain, breast, and squamous cell carcinomas. Inhibition of activated Stat3 leads to decreased proliferation and apoptosis of many cancer-derived cell lines, while the introduction of a constitutively activated form of Stat3 into immortalized human breast epithelial cells and rodent fibroblasts results in cellular transformation. Collectively, these data suggest a role for Stat3 in oncogenesis. A new study from Chan et al. (see related article beginning on page 720) is the first to demonstrate a requirement for Stat3 in de novo epithelial carcinogenesis in vivo. Using the two-step model of chemically induced skin carcinogenesis, the authors demonstrated that mice deficient in Stat3 were completely resistant to skin tumor development.

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Stat3 is required for the development of skin cancer

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Signal transducer and activator of transcription 3 (Stat3) is a transcription factor that is constitutively activated in a variety of human malignancies, including prostate, lung, brain, breast, and squamous cell carcinomas. Inhibition of activated Stat3 leads to decreased proliferation and apoptosis of many cancer-derived cell lines, while the introduction of a constitutively activated form of Stat3 into immortalized human breast epithelial cells and rodent fibroblasts results in cellular transformation. Collectively, these data suggest a role for Stat3 in oncogenesis. A new study from Chan et al. (see related article beginning on page 720) is the first to demonstrate a requirement for Stat3 in de novo epithelial carcinogenesis in vivo. Using the two-step model of chemically induced skin carcinogenesis, the authors demonstrated that mice deficient in Stat3 were completely resistant to skin tumor development.

Stat's (signal transducers and activators of transcription) are a family of latent transcription factors that are activated in response to many cytokines and growth factors. Stat activation is dependent upon tyrosine phosphorylation, which induces dimerization via reciprocal phosphotyrosine–src homology domain 2 (phosphotyrosine–SH2) interaction between two Stat molecules. Activated Stat's translocate to the nucleus where they bind to consensus promoter sequences of target genes and activate their transcription (1) (Figure 1). Many tyrosine kinases, including JAKs (Janus kinases), RTKs (receptor tyrosine kinases), and non-RTKs can phosphorylate Stat proteins. In normal cells, Stat tyrosine phosphorylation is transient, lasting from 30 minutes to several hours.

Nonstandard abbreviations used: DMBA, 7,12-dimethylbenz[a]anthracene; *Ha-ras*, Harvey rat sarcoma virus oncogene; JAK, Janus kinase; LRC, label-retaining cell; RTK, receptor tyrosine kinase; SH2, src homology domain 2; Stat3, signal transducer and activator of transcription 3; TPA, 12-*O*-tetradecanoylphorbol-13-acetate; *v-Ha-ras*, *Ha-ras* homolog.

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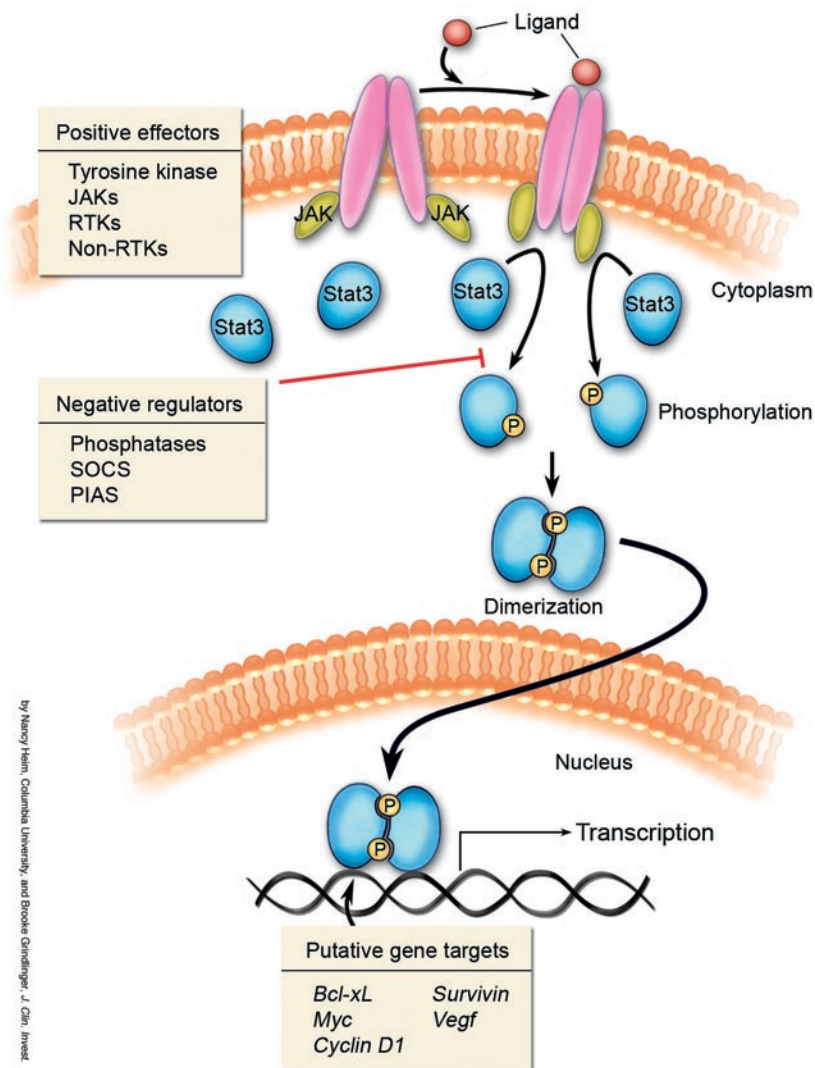
However, in numerous cancer-derived cell lines or in primary tumors, Stat proteins (in particular Stat3) are persistently tyrosine phosphorylated either as a consequence of deregulated positive effectors of Stat activation such as tyrosine kinases or negative regulators of Stat phosphorylation, e.g., phosphatases, suppressor of cytokine signaling, protein inhibitor of activated stats (2). Inhibition of Stat3 activity in tumor-derived cell lines by the introduction of antisense, small interfering RNA, dominant-negative Stat3 constructs, and/or blockade of tyrosine kinases has been associated with growth arrest and apoptosis (2). Furthermore, the introduction of a constitutively activated Stat3 molecule (Stat3C) into immortalized cell lines leads to transformation, indicating an oncogenic role for activated Stat3 (3, 4). A possible mechanism for transformation by activated Stat3 is the transcriptional upregulation of genes known to be involved in proliferation and apoptosis, including *Bcl-xL*, *c-Myc*, *cyclin D1*, *Vegf*, and *Survivin* (3, 5–8). In addition to its role as a transcription factor, phosphorylated Stat3 has been described in a recent report as a component of focal adhesions (sites of cell contact with the extracellular matrix) that may contribute

to the invasiveness of ovarian cancer cells (9). The in vivo role of Stat3 in tumorigenesis has not been addressed until now. In this issue of the *JCI*, Chan and colleagues demonstrate in two different murine models of skin tumor development that Stat3 is required for de novo tumorigenesis (10).

One of the best-established model systems for studying the mechanisms underlying the process of malignant transformation is the mouse skin model of multistage carcinogenesis (11, 12) (Figure 2). In this model, the process of skin tumor development can be subdivided into three different stages: initiation, promotion, and progression. Initiation is typically induced by the topical application of the carcinogen 7,12-dimethylbenz[a]anthracene (DMBA). Interestingly, in DMBA-treated epidermal cells, one usually finds mutations within the Harvey rat sarcoma virus oncogene (*Ha-ras*) gene. These mutations, however, are not sufficient to induce de novo transformation. Promotion of tumorigenesis is generated by the topical application of phorbol esters such as 12-*O*-tetradecanoylphorbol-13-acetate (TPA) to the skin, leading to epithelial cell proliferation with a concomitant increased expression of the ligand EGF as well as of cyclin D1, c-Jun, c-Fos, and c-Myc (13–16). TPA-treated mice form multiple benign papillomas within 10–20 weeks. Tumor progression is a spontaneous process resulting in the formation of malignant squamous carcinomas.

Stat3 prevents apoptosis in the initiation phase of skin tumorigenesis

K5Cre.Stat3^{fl/fl} transgenic mice, whose epidermal and follicular keratinocytes lack Stat3, are viable and develop normally.



by Nancy Heim, Columbia University, and Brooke Gruninger, J. Clin. Invest.

Figure 1

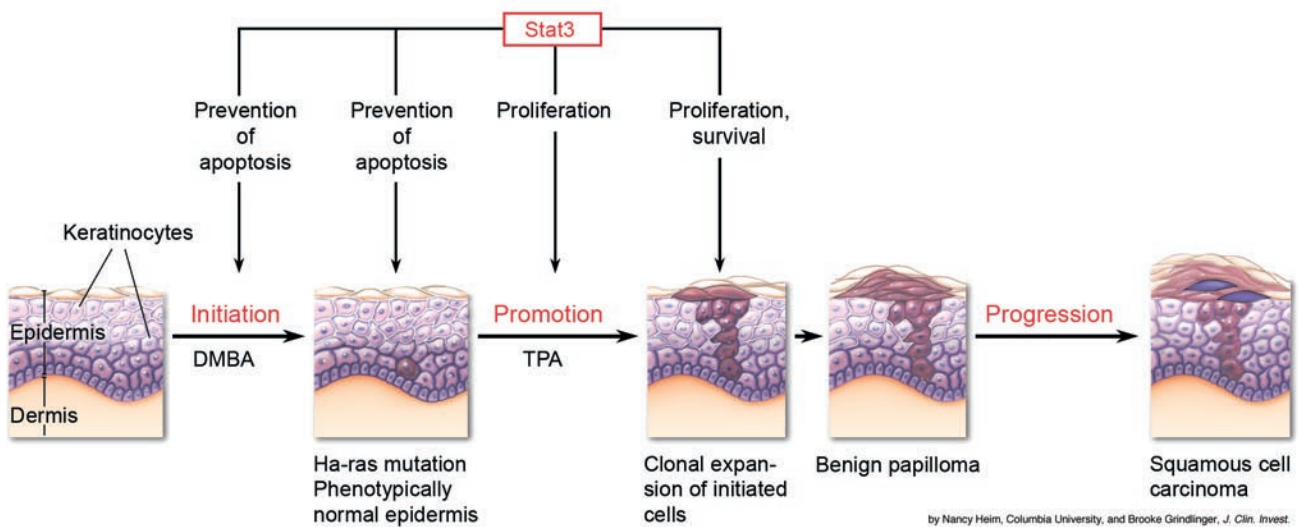
Model for Stat3 signaling. Stat3 is a transcription factor, which is activated in response to many cytokines and growth factors that bind to specific receptors. Upon ligand-receptor binding, Stat3 is recruited to the plasma membrane, where it becomes activated via phosphorylation of a tyrosine residue either directly by RTKs, such as the PDGF receptor and EGF receptor, or by non-RTKs, such as Src and JAK. Stat3 activation induces dimerization via reciprocal phosphotyrosine-SH2 interaction between two Stat3 molecules. The Stat3 dimers then translocate to the nucleus where they bind to consensus sequences on the promoter of target genes and activate their transcription. Stat3 activation is tightly regulated by different negative regulators of phosphorylation, such as phosphatases, suppressor of cytokine signaling, and protein inhibitor of activated Stats. In many cancer-derived cell lines and primary tumors Stat3 is constitutively activated either as a consequence of deregulated signaling from positive effectors (e.g., overexpression of growth factor receptors and their ligands) or by abnormal activity of negative effectors. SOCS, suppressor of cytokine signaling; PIAS, protein inhibitor of activated Stats.

of phosphorylated Stat3 present in these cells are sufficient to drive transcription of antiapoptotic genes such as *Bcl-xL*. Alternatively, nonphosphorylated Stat3 may be playing a role as a transcription factor as has been demonstrated for Stat1 (20). There are a few notable examples where relatively low levels of phosphorylated Stat3 are sufficient to mediate protection from growth arrest or apoptosis (21, 22). Thus, determination of the relative levels of phosphorylated Stat3 required to impart a phenotype is likely to be cell-type specific and remains an important objective. It has been shown that phosphorylated Stat1 levels are markedly enhanced in Stat3 null hepatocytes (23). Given that Stat1 activation has been implicated in promoting growth arrest as well as apoptosis, it would be of interest to determine whether the enhanced apoptosis observed in the Stat3 null keratinocytes correlates with increased levels of activated Stat1.

It is hypothesized that keratinocyte stem cells, which are located mostly within the bulge region of the hair follicle, are the target cells for two-stage carcinogenesis (24). Hair follicle stem cells have been identified within the label-retaining cells (LRCs), a population of cells that, following continuous administration of nucleotide analogs such as BrdU or [³H]thymidine, retains the label for a sustained period of time, indicating a very slow cycling frequency (25, 26).

However, they have defects in hair cycle processes as well as impaired wound healing, and they develop spontaneous ulcers with age. In vitro, keratinocytes derived from these mice had no defects in proliferation, but growth factor-dependent migration was markedly impaired in contrast to control keratinocytes (17). In the study described in this issue of the *JCI*, Chan et al. used K5Cre.Stat3^{fl/fl} transgenic mice to investigate the role of Stat3 in chemically induced carcinogenesis of the skin (10). The role of Stat3 in tumor initiation, the first step of chemically induced carcinogenesis, was addressed both in vitro, in DMBA-treated primary keratinocytes, and in vivo, by topical treatment with this mutagen. Stat3 mutant keratinocytes underwent enhanced apoptosis following DMBA treatment, compared to control keratinocytes. Expression of *Ha-ras* homolog (*v-Ha-ras*) into cultured primary

keratinocytes in vitro was used to generate initiated keratinocytes. Upon the introduction of a Stat3 decoy molecule (a high-affinity double-stranded DNA binding site for Stat3), the *v-Ha-ras*-initiated keratinocytes underwent apoptosis with a concomitant decrease in *Bcl-xL* levels. In general, inhibiting Stat3 function in cancer-derived cell lines containing abundant phosphorylated Stat3 leads to apoptosis or growth arrest. In contrast, cell lines which contain low or no levels of detectable tyrosine-phosphorylated Stat3 are relatively unaffected by Stat3 inhibitory therapies. It therefore remains unclear how Stat3 protects keratinocytes against DMBA-induced apoptosis, since DMBA does not induce tyrosine phosphorylation of Stat3 in primary keratinocytes, nor is it likely that *v-Ha-ras*-containing keratinocytes contain abundant levels of phosphorylated Stat3 (18, 19). Perhaps the low amounts



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Figure 2

Two-step model of chemically induced carcinogenesis and proposed role of Stat3 in preventing tumorigenesis. Initiation is achieved by topical application of the carcinogen DMBA, which binds to DNA and typically induces mutations in the *Ha-ras* gene. The result of this process is the formation of an initiated cell. During this stage Stat3 protects the keratinocytes and the initiated cells from DNA damage–induced apoptosis. Promotion involves repeated application of a noncarcinogenic promoter, TPA, leading to clonal expansion of the initiated cell. This process results in the formation of multiple benign papillomas. Stat3 is essential for the proliferation of the initiated cells after TPA treatment and for the maintenance of the preneoplastic lesions. Progression is a spontaneous process characterized by the accumulation of additional genetic changes (i.e., elevated expression of genes encoding *Ha-ras* and cyclin D1; loss of functional p53), which accelerate the conversion of benign papillomas to malignant carcinomas. The pink cells represent initiated cells bearing *Ha-ras* mutations; the purple cells represent cells that have acquired multiple genetic changes.

Chan et al. (10) observed that the majority of the Stat3-deficient keratinocytes undergoing apoptosis after exposure to DMBA were located primarily within the bulge region of the hair follicle in an area adjacent to the LRC population. The authors suggest that the DMBA-sensitive cells may be keratinocyte stem cells, given their proximity to the LRCs. However, given the complete lack of overlap between the LRCs and the apoptotic cells, the cell type most sensitive to DMBA-induced apoptosis remains to be identified. The work of Chan et al. generates interesting questions regarding the mechanism(s) by which Stat3 affords protection against apoptosis and the determination of which cell type(s) are most sensitive to the loss of Stat3.

Stat3 is required for keratinocyte proliferation during the promotion stage of chemically induced carcinogenesis

Tumor promotion, the second stage of chemically induced skin carcinogenesis, occurs following the topical application of TPA, leading to the proliferation of epidermal cells and the subsequent formation of benign papillomas in wild-type animals. Perhaps the most important observation made by Chan et al. (10) is that mice defi-

cient in Stat3 were completely resistant to skin tumor development. Given that DMBA treatment resulted in keratinocyte apoptosis in the K5Cre.Stat3^{fl/fl} animals, there was undoubtedly a decreased number of keratinocytes capable of proliferating in response to TPA. Nevertheless, the role of TPA alone (in the absence of the initiation stage) was assessed in vivo by applying TPA topically. Marked epidermal hyperproliferation and BrdU uptake was observed in wild-type animals. In contrast, significantly less proliferation was seen in the K5Cre.Stat3^{fl/fl} animals. TPA induced the tyrosine phosphorylation of Stat3, perhaps in part through increased production of EGF leading to activation of the EGFR (13). An increase in some of the known putative Stat3 targets, including *cyclin D1* and *c-Myc*, was observed in the skin of wild-type animals while in the K5Cre.Stat3^{fl/fl} animals a delay in the increased level of these proteins was seen. These data demonstrate that upregulation of these genes is partly dependent upon Stat3. The requirement of Stat3 in tumor promotion was also examined in transgenic animals expressing v-Ha-ras targeted to keratinocytes. Topical TPA treatment of these animals led to benign papilloma formation, which was abrogated upon the

introduction of a Stat3 decoy molecule. The use of a Stat3 decoy was first described as an inhibitor of tubular morphogenesis in cells grown in culture (27). It was subsequently shown to inhibit growth of squamous cell carcinoma–derived cell lines in vitro and more recently in vivo (28) (J. Grandis, personal communication).

The use of this molecule in vivo is indeed novel, as is the manner in which the Stat3 decoy was administered (10). The Stat3 decoy was applied topically to the mice on a weekly basis. It is remarkable that the DNA was absorbed in sufficient quantities to exert a profound inhibitory effect on papilloma formation. Intralesional injection of the Stat3 decoy was also effective in decreasing the size of existing papillomas. It would be of interest to know whether an increase in apoptosis was observed in the Stat3 decoy–injected lesions, as has been described for preformed B16 melanoma tumors injected with a dominant negative Stat3 construct (29).

Decoy molecules against Stat3 may indeed be effective therapy for cutaneous or locally advanced cancers, including skin cancers, head and neck tumors, and metastatic breast cancers involving the skin. One potential caveat with the Stat3 decoy relates to the presumed concomitant inhi-



bition of Stat1 activity. The decoy should bind equally well to the Stat3 and Stat1 dimers. Given the evidence that Stat1 may function as a tumor suppressor as well as a promoter of apoptosis, the sequestering of the Stat1 dimer may oppose the tumoricidal effects of sequestering Stat3 (30–32). Nevertheless, this model system of skin tumorigenesis is an ideal one for developing and testing anti-Stat3 drugs for the prevention and treatment of skin cancer.

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